# Preparation and Characterization of Imazalil / **b**-Cyclodextrin Inclusion Complex by Supercritical Carbon Dioxide and <sup>13</sup>C Cp-Mas and <sup>1</sup>H NMR Spectroscopy: Preliminary Results

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## ABSTRACT

Imazalil, a selected fungicide, was inclused into  $\beta$ -cyclodextrin using supercritical carbon dioxide. The best experimental conditions were determined to prepare the insertion complex, which was investigated by means of <sup>1</sup>H NMR in aqueous solution and <sup>13</sup>C CP-MAS NMR in the solid state. A comparative analysis of the spectra of the free  $\beta$ CD, IMZ and  $\beta$ CD-IMZ gave information on structural properties of the  $\beta$ CD-IMZ complex.

## **INTRODUCTION**

Imazalil (IMZ), one of the few fungicides permitted in the post-harvest treatment of fruits [1-3], is poorly soluble in water (0.018g/100mL), that reduces notably its antifungal activity in the agricultural applications. To improve the solubility in water, IMZ was microencapsulated in  $\beta$ -cyclodextrin ( $\beta$ CD) by using supercritical carbon dioxide (SCF).

Generally, to prepare  $\beta$ CD-drug inclusion complexes several preparation methods in solution are available [4-6]. However, many of these methods are time-consuming and need multistage processing, involving evaporation of large volumes of solvent, which often occurs in the complex as a residue.

SCF approch permits to avoid such disadvantages and, in particular, to improve the purity of the  $\beta$ CD-IMZ complex, being moreover a well known environmental sustainable method [7-10].

Information on the stoichiometry and structure of this inclusion complex was obtained using <sup>1</sup>H NMR in aqueous solution and <sup>13</sup>C CP-MAS.

# MATERIAL AND METHODS

#### Materials

 $\beta$ -CD (CAVAMAX <sup>®</sup>7 PHARMA) was obtained from Wacker – Chemie Italia SpA and was used as received; CO<sub>2</sub> (purity 99 %) was supplied by SIO (Società Italiana Ossigeno, Cagliari, Italy), while IMZ (97%) was purchased from Dr. Ehrenstorfer (Augsburg, Germany) and used without further purification.

## SCF Apparatus

Supercritical  $CO_2$  inclusions were performed in a laboratory apparatus equipped with a 400 cm<sup>3</sup> autoclave, thermostated and supercritical  $CO_2$  pressurized by a high pressure diaphragm pump (Lewa, model EL 1) with a maximum capacity of 6 kg/h, pumped liquid  $CO_2$  at the desiderate pressure. Temperatures and pressures along the extraction apparatus were measured by thermocouple and Bourdon-tube test gauges, respectively. Pressure was regulated by high pressure valves under manual control [11].

The apparatus has been used in the batch mode. The autoclave was charged with about 20 g of  $\beta$ CD and with the guest compound at a molar ratio of 1:2.5. Then, thermostated and pressurized with CO<sub>2</sub> at the fixed constant value for the prefixed time of contact. Then a rapid drop of pressure permitted to vaporized CO<sub>2</sub> and to separate the solid deprived of solvent.

# Process parameters of the SCF preparations

The following equilibrium is obtained when one drug molecule (D) forms a 1:1 complex with  $\beta$ CD:

 $\begin{array}{l} \beta CD + D \iff \beta CD\text{-}D \qquad (K_{1:1} \ ) \\ \text{where } \beta CD\text{-}D \ \text{is the insertion complex and } K_{1:1} \ \text{is the apparent constant of the complex. The complexation efficiency (CE) can be defined as the } [\beta CD\text{-}D]/[\beta CD] \ \text{molar ratio in the solution. If the solution is saturated with the drug, the concentration of the free drug ([D]) will be equal to the intrinsic solubility of the drug (S_0).} \end{array}$ 

# NMR experiments.

<sup>1</sup>H NMR spectra of the  $\beta$ -CD/IMZ complex in D<sub>2</sub>O were recorded in 5 mm tubes on a Varian VXR-300 spectrometer at T = 300 K. Residual water signal was suppressed by presaturation with low irradiation power.

Cross-polarization magic-angle spinning (CPMAS)  $^{13}$ C NMR spectra were recorded on a Varian 400 Unity Inova spectrometer at a resonance frequency of 100.57 MHz. A matched Hartmann-Hann condition was established at the spin-lock field of 38 KHz. The rotor was spun at a rate of 4 KHz and, working at room temperature, a contact time of 500 µs was applied to obtain polarization transfer. A recycle delay of 5 s and the 90 degree pulse of 5.8 µs were used. Chemical shifts of CPMAS spectra were originally obtained with respect to the methylene carbon resonance of solid adamantane, 38.3 ppm with respect to Me<sub>4</sub>Si, measured before each measurement. Experiments were conducted on 160-180 mg samples of cyclodextrin packed into a 7 mm ZrO<sub>2</sub> rotor.

## **RESULTS AND DISCUSSION**

### Process parameters of the SCF preparations

To improve the efficiency in the complex preparation of the  $\beta$ CD-IMZ insertion complex by SCF, we have tested the process thermodynamic parameters, such as temperature and pressure, in order to increase K<sub>1:1</sub> and S<sub>0</sub> in supercritical CO<sub>2</sub> medium. We have also monitored the influence of the contact time IMZ- $\beta$ CD-CO<sub>2</sub> on the complexation efficiency, using different temperatures (55, 65, 75, 85 °C) at P = 150, 200 bar, different pressures (130, 150, 170, 200 bar) at T = 65 °C and different contact times (2, 4, 6 h) at T = 65 °C and P = 150 and 200 bar (Table I). The data reported in Table I show that the complexation efficiency increases as a function of the contact time in both the experiments at 150 bar and at 200 bar. As shown, a contact time of 6 hours is necessary to achieve the best distribution equilibrium between the supercritical phase and the solid phase. In addition, the complexation efficiency increases as the pressure increases (from 130 bar to 200 bar at 65 °C) due to an increases solubility of IMZ in supercritical CO<sub>2</sub>, since the density and the solvent power of CO<sub>2</sub> increases as a function of the pressure.

Table I shows that the complexation efficiency increases as the temperature increases (from 55 to 85 °C) at 150 bar, while at 200 bar it decreases at the increasing of the temperature (from 65 to 85 °C). This result can be explained considering that the temperature has two opposite effects. In fact, on the one hand, the increase of temperature causes an increase of the vapour tension of IMZ, thus enhancing its solubility in supercritical  $CO_2$ , on the other hand, too high temperatures cause the decrease of density of supercritical  $CO_2$  thus decreasing its solvent power [8]. It stands to reason that the crossing point (75 °C) is very

close to the temperature value (65 °C) where a reversal of behaviour of IMZ solubility in supercritical  $CO_2$  was observed.

In the light of these findings, evidence is given that P = 200 bar, T = 65 °C and contact time = 6h are the best conditions to obtain a good efficiency in the complex formation (complexation efficiency = 1.0).

Although an excess of guest molecule was used in all the preparations by SCF only equimolar guest-host ratio was achieved.

# <sup>13</sup>C CP-MAS and <sup>1</sup>H *NMR* characterization

Evidence of the formation of the  $\beta$ CD-IMZ complex was given by <sup>1</sup>H NMR spectroscopy in aqueous solution. In Figure 1 a comparison between the <sup>1</sup>H NMR spectra of the free and complexed  $\beta$ CD is shown. It is easy to see that in the spectrum of the  $\beta$ CD-IMZ complex the IMZ signals also appear, despite the very low solubility of IMZ in water, clearly indicating that the formation of the inclusion complex occurred. The assignment of the resonances was made following previous NMR investigations [4]. As shown, the resonances of the  $\beta$ CD in the inclusion complex exhibit significant changes in chemical shifts and linewidths. In particular, relevant highfield shifts (about 0.2 ppm) were detected for H<sub>3</sub> and H<sub>5</sub> protons, which point into the hydrophobic cavity of the  $\beta$ CD, in agreement with the results obtained for the inclusion complex prepared in water [4].

A stoichiometry of approximately 1:1 was determined by the integration of the host and guest signals in the <sup>1</sup>H spectrum of the  $\beta$ CD-IMZ [4].

Figure 2 shows the solid-state <sup>13</sup>C CP-MAS NMR spectra of the free  $\beta$ CD and IMZ. The assignment of the resonances was made following previous NMR investigations [4] and for IMZ was obtained from the analysis of the coupled and decoupled spectra in CDCl<sub>3</sub> (see Table II).

As it can be observed, the spectrum of  $\beta$ CD has a relatively poor resolution, due to the partial hydration of the sample, however, multiple resonances appear mainly for the C<sub>2,3,5</sub>, C<sub>4</sub> and C<sub>6</sub> atoms of the glucopyranose unit, that indicates the coexistence of different structural arrangements [12]. The spectrum of the inclusion complex show that all of the major signals of  $\beta$ CD shift and change in intensity compared to their free components, and a marked broadening of the signals of the guest molecule occurs.

It is note worthy that the parameters responsible for these changes are differently affected. In fact, each set of multiple resonances of carbons of the glucose units tends to converge to a single peak, thus suggesting that in the complex the glucose units adopts a more symmetrical conformation.

The analysis of the spin-lattice relaxation times in the rotating frame,  $T_{1\rho}\ell^{l}H$ ), and of the <sup>129</sup>Xe NMR spectra is now in progress to obtain information on the dynamics and the hydrophobic character of the  $\beta$ CD-IMZ complex .

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**Table 1:** Experimental parameters used to prepare  $\beta$ CD-IMZ inclusion complex: T, temperature (°C); P, pressure (bar); t, time of contact (h); R, starting stoichiometry ratio of free  $\beta$ CD and IMZ; CE, complexation efficiency of the complex,  $\beta$ CD-IMZ equivalents ratio.

T/°C	P/bar	t/h	R	CE	
65	130	6	2.5	0.73	
65	150	6	2.6	0.80	
65	170	6	2.6	0.87	
65	200	6	2.6	0.96	
55	150	6	2.7	0.74	
65	150	6	2.6	0.80	
75	150	6	2.7	0.88	
75	150	6	2.6	0.86	
75	150	6	2.8	0.84	
85	150	6	2.6	0.92	
65	200	6	2.6	0.96	
05 75	200	6	2.0	0.90	
85	200	6	2.0	0.00	
85	200	6	2.5	0.69	
65	150	2	2.7	0.67	
03 65	150	2	2.7	0.07	
00 (5	150	4	2.7	0.74	
65	150	6	2.6	0.80	
65	200	2	2.7	0.83	
65	200	4	2.6	0.90	
65	200	6	2.6	0.96	

	Carbon type	<b>d</b> (ppm)		
		Free	insertion complex	-
	$C_1$	103.8	104.1	_
		103.0	103.0	
	$C_4$	84.9	84.3	
		84.0	83.1	
		82.8	82.1	
		82.0	80.9	
bCD		79.0	79.1	
		76 9	76 9	
	$C_2/C_3/C_5$	70.8	70.8	
		73.3	13.9	
		74.3		
		73.5		
	C	61 1	61 2	
	$C_6$	04.4 62.5	04.5 62.4	
		60.5	61.0	
	N-CH=N	139.1	138.5	17
IMZ	Cl-C (ar)	136.5	135.8	16
	$-C_3H=(ar)$	131.2	130.0	
	$-C_{5,6}H=(ar)$	129.4	128.7	15 - 13 - 12
	$N-C_{11}H=$	125.7	124.7	
	$N-C_{10}H=$	120.4	119.6	$1 \downarrow 9_{\rm N} / 11$
	O-CH-	76.1	nd	5 7 <u>8</u> 10
	O-CH <sub>2</sub> -	71.7	nd	
	N-CH <sub>2</sub> -	52.8	52.5	Cl 4 2 Cl

**Table 1:** <sup>13</sup>C chemical shifts,  $\delta$  (ppm), of  $\beta$ CD, IMZ and  $\beta$ CD-IMZ.



Figure 1: H NMR spectra of  $\beta$ CD-IMZ (a, b) and  $\beta$ CD (c) in aqueous solution.



**Figure 2:** <sup>13</sup>C CP-MAS NMR spectra of  $\beta$ CD (A), IMZ (B) and  $\beta$ CD-IMZ (C). \* Spinning sidebands.